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**INVITED ABSTRACTS OF THE 4TH EUROPEAN SYMPOSIUM  
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**Reductive activation of methanol: 5-hydroxybenzimidazolylcobamide methyltransferase from Methanol sarcina barkeri**

The conversion by Methanosarcina barkeri of methanol into methylcoenzyme M (2-methyl-thioethanesulfonate,  $\text{CH}_3\text{S-CoM}$ ), the substrate of the methane-forming reaction, is catalyzed by a complex enzyme system including methanol: 5-hydroxybenzimidazolylcobamide methyltransferase ( $\text{MT}_1$ ) (Fig. 35) [1].  $\text{MT}_1$  is a 135 kDa protein that harbours as its prosthetic groups two molecules 5-hydroxybenzimidazolylcobamide ( $\text{B}_{12}\text{HBI}$ ). In the reaction cycle  $\text{MT}_1$  accepts the methyl group from methanol and binds it as methyl- $\text{B}_{12}\text{HBI}$ . Subsequently, the methyl group is transferred by a second methyltransferase ( $\text{MT}_2$ ) to coenzyme M (2-mercaptoethanesulfonate, Coenzyme M). In the in vitro reaction an additional set of enzymes and coenzymes are required, notably ATP (in catalytic amounts), Methyltransferase Activating Protein (MAP), hydrogen, hydrogenase, and ferredoxin [1]. The latter play a role in the (re)activation of  $\text{MT}_1$ , when the enzyme has become inactive as the result of the oxidation of the catalytic competent  $\text{Co(I)B}_{12}\text{HBI}$ .

Inactive  $\text{MT}_1$  contained  $\text{B}_{12}$  as the  $\text{Co(III)}$  and  $\text{Co(II)}$  species and incubation with hydrogen, hydrogenase and ferredoxin did not allow a reduction beyond the  $\text{Co(II)}$  level. EPR experiments demonstrated the 5-hydroxybenzimidazole base to be coordinated to the cobalt atom ('base-on'). When both ATP and MAP were supplied,  $\text{Co(II)-B}_{12}\text{HBI}$  was transformed into the 'base-off' state. This has an important implication, since studies with isolated  $\text{B}_{12}\text{HBI}$  showed that the midpoint reduction potential of 'base-off'  $\text{Co(II)-B}_{12}\text{HBI}$  to  $\text{Co(I)-B}_{12}\text{HBI}$  ( $E_0 = -500$  mV) is about 100 mV more positive as compared to the same reduction of 'base-on'  $\text{Co(II)B}_{12}\text{HBI}$  [2]. Yet, the potential still may be too low for reduction with hydrogen ( $E'^{\text{sub }0}(2\text{H}^+/\text{H}_2) = -414$  mV). However, when methanol was also present in the reactivation mixture, the corrinoids in  $\text{MT}_1$  were methylated and hereafter the enzyme had become fully active. This suggests that methanol is required to trap  $\text{Co(I)}$  out of the 'base-off'-Co

(II)/Co(I) equilibrium. By using purified MAP it was found that the enzyme was phosphorylated by ATP. MAP-phosphate substituted for ATP in the activation of  $MT_1$  indicating that the phosphorylated enzyme catalyzed the critical step in the activation process: the conformational change of 'base-on' into 'base-off' Co(II)- $B_{12}$ HBI.

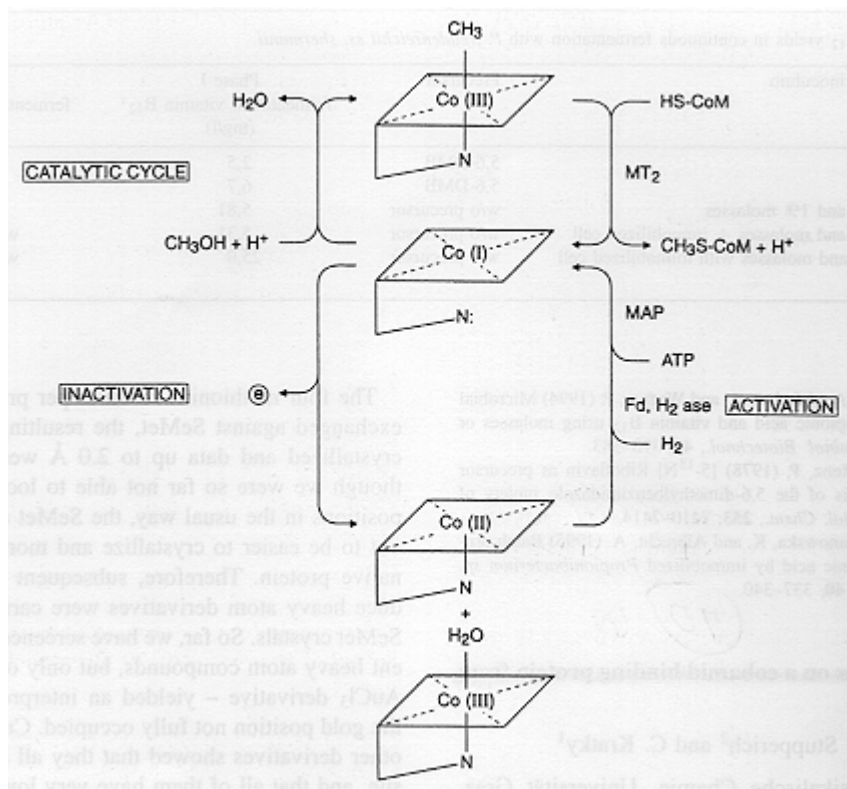


Fig. 35. The methanol: HS-CoM methyltransferase system. Methanol: 5-hydroxybenzimidazolylcobamide methyltransferase ( $MT_1$ ) is schematically represented by its corrinoid prosthetic group;  $MT_2$ , methylcobamide: HS-CoM methyltransferase; MAP, Methyltransferase Activating protein;  $H_2ase$ , hydrogenase; Fd, ferredoxin.

### References

- [1.] Daas, P.J.H., Gerrits, K.A.A., Keltjens, J.T., van der Drift, C. and Vogels, G.D. (1993) *J. Bacteriol.*, 175, 1278-1283.
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